

## 2 D-NMR of Natural Products, Part V<sup>1</sup> Structure Elucidation and Complete <sup>1</sup>H- and <sup>13</sup>C-Assignment of Resin Acid Derivatives

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Application of 2 D-NMR-techniques including heteronuclear 2 D-*J*-resolved spectroscopy and <sup>1</sup>H—<sup>13</sup>C-2 D-shift correlation is used to assign the <sup>1</sup>H- and <sup>13</sup>C-resonances of resin acid derivatives.

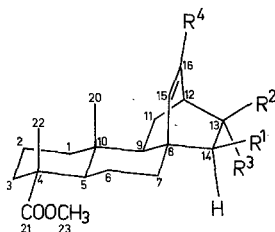
(Keywords: 2 D-NMR; 2 D-*J*-Resolved NMR spectroscopy; <sup>1</sup>H—<sup>13</sup>C-Shift correlation; Diterpene acids)

*2 D-NMR von Naturprodukten, 5. Mitt.: Strukturaufklärung und komplette <sup>1</sup>H- und <sup>13</sup>C-Zuordnung von Harzsäure-Derivaten*

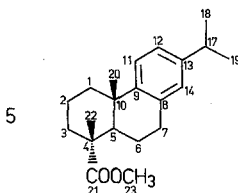
2 D-NMR-Methoden (2 D-*J*-resolved-Spektren, <sup>1</sup>H—<sup>13</sup>C-Shiftkorrelations-spektren) erlauben die Zuordnung der <sup>1</sup>H- und <sup>13</sup>C-Signale von Harzsäurederivaten.

### Introduction

During our attempts to use resin acids as precursors for the synthesis of steroids we have studied various *Diels-Alder*-adducts of levopimaric acid<sup>2,3</sup>. One of the main problems in the course of this work is the characterization of reaction products by NMR. Even using highfield NMR-spectrometers, the analysis of the proton spectra of complex natural compounds like resin acids is not possible with conventional methods. In these cases 2 D-NMR-techniques have proved to be very useful for obtaining additional information<sup>4,5</sup>. We have used several of these modern techniques simultaneously to get an unambiguous assignment of the <sup>1</sup>H- and <sup>13</sup>C-resonances of compounds **2**, **3** and **4**.



	R <sup>1</sup>	R <sup>2</sup>	R <sup>3</sup>	R <sup>4</sup>
1		$\begin{array}{c} \text{C} \\ \parallel \\ \text{O} \end{array} \text{O} \text{C} \begin{array}{c} \parallel \\ \text{O} \end{array}$	-H	$-\text{CH}(\text{CH}_3)_2$
2		$\begin{array}{c} \text{C} \\ \parallel \\ \text{O} \end{array} \text{O} \text{C} \begin{array}{c} \parallel \\ \text{O} \end{array}$	-H	$\begin{array}{c} \text{CH}_2 \\ \parallel \\ \text{C} \\ \parallel \\ \text{CH}_3 \end{array}$
3	$-\text{COOCH}_3$	-H	$-\text{COOCH}_3$	$-\text{CH}(\text{CH}_3)_2$
4	$-\text{COOCH}_3$	-H	$-\text{COOCH}_3$	$-\text{CBr}(\text{CH}_2\text{Br})_2$



For the sake of clarity we have used the same numbering system for compounds **1** to **5**.

### Results and Discussion

The assignment of the <sup>1</sup>H- and <sup>13</sup>C-resonances of compounds **2–4** is given in Tables 1 and 2, respectively, and will be discussed in detail. For the sake of clarity the shifts of compounds **1** and **5** are also listed in the tables<sup>5,6</sup>. The assignment of **5** was checked by 2D-NMR-experiments and is in full agreement with literature.

#### Compound 2

The assignment of the carbons 15, 18 and 21 is possible by shift arguments and multiplicity information from a *J*-modulated <sup>13</sup>C-spectrum. The anhydride carbons and the two remaining singulets at 137.9 and 140.6 ppm could not be assigned unambiguously.

The aliphatic region of the <sup>13</sup>C-spectrum consists of 18 well separated lines. The signal at 51.0 ppm is due to the ester-methyl group. The methyl group attached to C-17 can be easily detected in the SFORD-

Table 1.  $^{13}\text{C}$ -Chemical shifts of compounds **1**–**5** in  $\text{C}_6\text{D}_6$ ;  $\delta$ -values are in ppm from TMS ( $\pm 0.05$ ); <sup>a,b,c</sup> shift values may be interchanged

	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>
1	38.1	37.9	37.8	37.6	38.4
2	17.3	17.3	17.4	17.4	18.9
3	37.2	37.2	37.1	37.0	37.1
4	47.2	47.2	47.4	47.2	47.8
5	49.6	49.5	49.7	49.4	45.4
6	22.0	22.0	22.3	22.1	22.2
7	35.2	34.9	35.3	34.6	30.4
8	40.4	41.0	41.7	42.2	147.3
9	53.1	53.2	54.8	55.1	134.8
10	37.7	37.6	38.0	37.8	37.2
11	27.2	26.3	23.9	24.5	124.5
12	35.8	33.0	36.3	36.8	124.2
13	45.7	45.6	49.1	48.8	145.8
14	53.0	52.8	54.5	55.5	127.3
15	125.4	129.1	125.0	134.7	—
16	148.1	141.9 <sup>a</sup>	148.2	138.9	—
17	33.0	139.3 <sup>a</sup>	33.1	67.0	34.0
18	20.1 <sup>a</sup>	112.9	20.6	38.9 <sup>a</sup>	24.3
19	20.7 <sup>a</sup>	19.7	20.6	37.4 <sup>a</sup>	24.3
20	17.0	17.0	17.1	17.2	16.9
21	178.4	178.4	178.5	178.3	178.3
22	15.5	15.2	16.3	16.1	25.3
23	51.6	51.6	50.9 <sup>a</sup>	51.1 <sup>b</sup>	51.5
24	172.6 <sup>b</sup>	171.9 <sup>b</sup>	174.2 <sup>b</sup>	173.2 <sup>c</sup>	—
25	170.8 <sup>b</sup>	170.8 <sup>b</sup>	173.8 <sup>b</sup>	173.0 <sup>c</sup>	—
26	—	—	51.5 <sup>a</sup>	51.5 <sup>b</sup>	—
27	—	—	51.5 <sup>a</sup>	51.8 <sup>b</sup>	—

spectrum by its large residual splitting. The proton signal of Me-22 is shifted highfield by the anisotropy of the C-15/C-16-double bond, thus yielding the assignment of C-20 and C-22 via the 2D- $^1\text{H}$ - $^{13}\text{C}$ -shift correlation.

In the  $^1\text{H}$ -spectrum the protons H-7 eq, H-12, H-13 and H-14 show characteristic splitting patterns and are well separated. The shift correlation spectrum therefore allows the assignment of C-7, C-12, C-13 and C-14. C-7 gives in turn the resonance position of H-7 ax.

The assignment of the remaining triplets in the lowfield region of the  $^{13}\text{C}$ -spectrum (C-1 and C-3) is possible because of the characteristic highfield shift of H-1 ax<sup>5</sup>. The assignment of the remaining  $^{13}\text{C}$ -signals was done by comparing spectra of **1** and **2**, yielding the shift values of all protons by the shift correlation experiment.

Compound **3**

The  $^1\text{H}$ -spectrum shows an isopropyl group and two further aliphatic methyl groups. The corresponding  $^{13}\text{C}$ -resonances can be detected via the shift correlation experiment using the same arguments for the assignment of H-22 and H-20 as above.

Table 2.  $^1\text{H}$ -Chemical shifts of compounds **1-5** in  $\text{C}_6\text{D}_6$ ;  $\delta$ -values are in ppm from TMS ( $\pm 0.01$ ); <sup>a,b,c</sup> shift values may be interchanged

	<b>1</b>	<b>2</b>	<b>3</b>	<b>4</b>	<b>5</b>
1 eq	1.03	0.97	1.14	1.12	2.12
1 ax	0.63	0.61	0.68	0.63	1.38
2	1.18-1.25	1.17-1.26	~ 1.2	~ 1.2	~ 1.4
3 eq	1.50	1.52	1.47	1.46	1.58
3 ax	1.77	1.74	1.80	1.68	1.83
5	1.71	1.75	1.81	1.75	2.32
6	1.18-1.25	1.17-1.26	~ 1.3	~ 1.2	1.40/1.73
7 eq	2.54	2.54	1.78	1.91	2.74
7 ax	1.53	1.50	1.71	1.53	2.74
9	0.77	0.96	1.35	1.41	—
11 eq	0.73	0.73	0.93	1.12	—
11 ax	0.98	0.80	1.67	1.62	7.13
12	2.74	3.28	2.92	3.14	7.02
13	2.30	2.35	2.87	3.05	—
14	1.53	1.97	3.06	3.11	6.86
15	5.23	5.56	5.48	5.72	—
17	2.08	—	2.30	—	2.74
18	0.85 <sup>a</sup>	4.88/5.19	1.07	3.79/3.93 <sup>b</sup>	1.19
19	0.86 <sup>a</sup>	1.58	1.07	3.90/4.05 <sup>b</sup>	1.19
20	1.15	1.13	1.20	1.17	1.27
22	0.26	0.22	0.46	0.45	1.10
23	3.40	3.40	3.40	3.23 <sup>c</sup>	3.35
26	—	—	3.51	3.38 <sup>c</sup>	—
27	—	—	3.51	3.34 <sup>c</sup>	—

H-12, H-13 and H-14 are separated from all other  $^1\text{H}$ -resonances, thus giving easily the corresponding carbon signals. Ring opening of the anhydride causes a highfield shift of H-7eq while C-7 remains unaffected. Irradiation at H-12 gives the resonance position of the protons at C-11, leading to an assignment of the carbon atom itself. C-1 and C-3 can be differentiated by the corresponding  $^1\text{H}$ -chemical shifts. The assignment of the residual resonances was done in analogy to the other compounds.

## Compound 4

The  $^{13}\text{C}$ -spectrum of **4** shows several groups of resonances with nearly identical shift values. Therefore  $^{13}\text{C}$ -multiplicity assignment was performed by a  $^{13}\text{C}$ - $^1\text{H}$ - $J$ -resolved experiment (Fig. 1).

The  $^1\text{H}$ -spectrum of **4** exhibits two overlapping AB-systems near 4 ppm which correspond to H-18 and H-19 giving the shifts of C-18 and C-19. The assignment of the carbons and protons at positions 12, 13 and

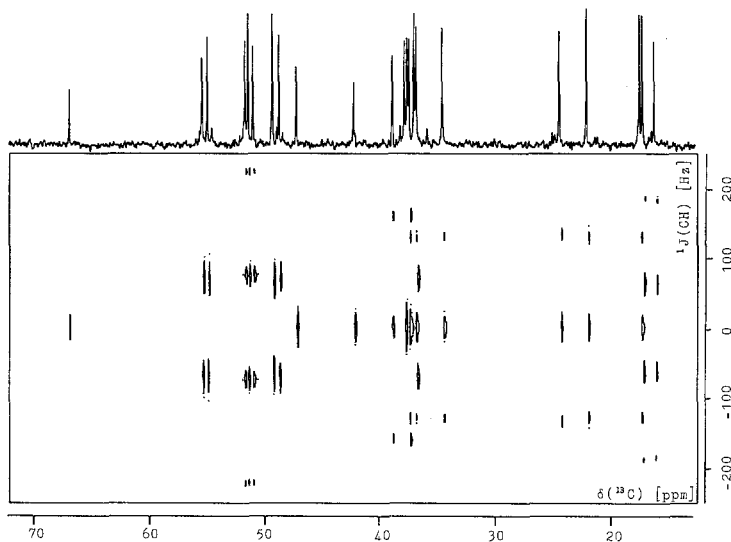


Fig. 1.  $^{13}\text{C}$ - $^1\text{H}$ - $J$ -resolved NMR spectrum of **4** in  $\text{C}_6\text{D}_6$

14 was done in the usual manner. The  $^1\text{H}$ -resonance of H-7 eq is well separated giving C-7 and in turn H-7 ax via the shift correlation spectrum (Fig. 2). Me-22 appears in the  $^1\text{H}$ -spectrum at highest field leading to the assignment of C-22. Discrimination of C-1 and C-3 is possible by the highfield shift of H-1 ax leading to the assignment of C-1 and C-3. The remaining three triplets (C-2, C-6 and C-11) could be assigned by comparison of **4** with the other compounds (see Table 1). The low-field singulett at 67.0 ppm belongs to C-17, the two doublets for C-5 and C-9 are found at 49.4 ppm and 55.1 ppm, respectively.

Comparing the  $^{13}\text{C}$ -shifts of compounds **1-4** one finds that the shifts of carbons 9, 11, 13 and 14 are sensitive to the orientation of the carbonyl groups attached to the C-ring. Introduction of a C=C-double bond in position 17 induces a lowfield shift for C-15 and a highfield shift for C-

16<sup>7</sup>. The structure determination of compound **4** was mainly based on the results of the 2D-NMR-shift correlation. The influence of the Br-substitution on the <sup>13</sup>C-shifts can be seen by comparing the shift values of **3** and **4**. Only the two olefinic carbons are shifted considerably (C-15: 10 ppm to low field, C-16: 10 ppm to high field). This is probably due to

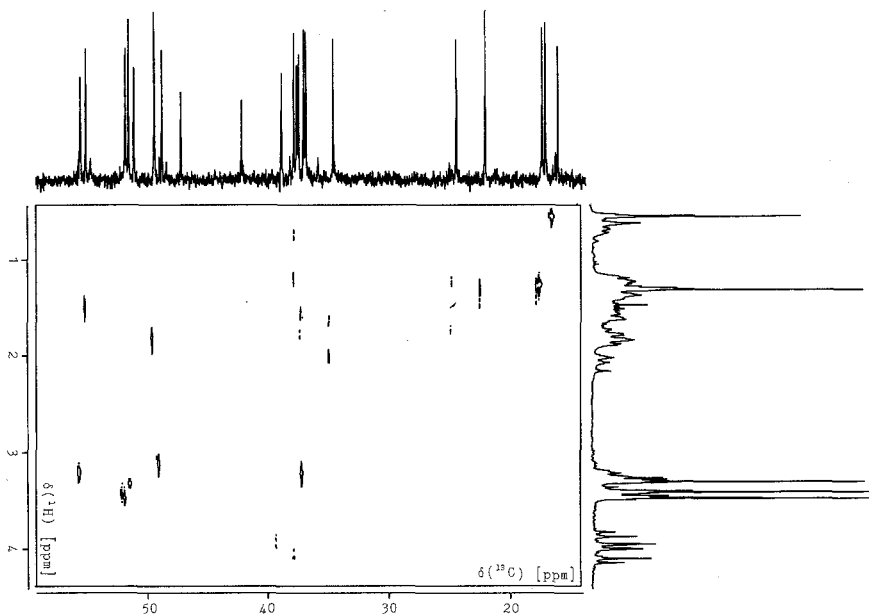


Fig. 2. <sup>13</sup>C—<sup>1</sup>H-shift correlation spectrum of **4** in C<sub>6</sub>D<sub>6</sub>

electronic and steric interactions with the Br-atoms. The <sup>1</sup>H-shifts reveal no big influence of the Br-substituents on the surrounding protons. Only the equatorial hydrogens attached to the C-ring are shifted slightly to lower field in compound **4**.

Finally we have computed theoretical shift values with a computer program from a <sup>13</sup>C-data bank<sup>8</sup>. Theoretical and experimental shift values have been in excellent agreement.

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## Experimental

The syntheses of compounds **1**–**5** have been described elsewhere<sup>3,9,10</sup>.

All NMR-measurements were performed on a Bruker WM 250 NMR spectrometer equipped with an 80 K ASPECT-2000 computer running the DISNMRP-program<sup>11</sup>. Sample tubes of 5 mm diameter have been used. The deuterium resonance of the solvent provided the field-frequency-lock.

Typical parameters were:

<sup>1</sup>H

$SW = 2\,500$  Hz; digital resolution: 0.3 Hz;  $PW = 1\ \mu\text{s}$  ( $15^\circ$ );  $NS = 32$ – $64$ ;  $T = 298$  K; concentration: approx. 0.02 *M*.

<sup>13</sup>C

$SW = 16\,000$  Hz; digital resolution: 1 Hz;  $PW = 5\ \mu\text{s}$  ( $30^\circ$ );  $NS = 500$ – $2\,000$ ;  $T = 303$  K; concentration: approx. 0.4 *M*.

### *J*-Modulated <sup>13</sup>C-Spin-Echo

Pulse sequence: D  $1-\pi/2-\tau(\text{DO})-\pi-\tau(\text{BB})$ -FID;  $SW = 16\,000$  Hz;  $NS = 500$ – $2\,000$ ;  $PW(90^\circ) = 16\ \mu\text{s}$ ;  $T = 303$  K;  $\tau = 7.5$  ms for  $J = 133$  Hz; recycle delay: 2.5 s.

### Heteronuclear 2 D-*J*-Resolved Spectrum

Pulse sequence: D  $1-\pi/2(\text{PH } 1, \text{BB})-t_1/2-\pi(\text{PH } 2, \text{DO})-t_1/2$ -FID (BB);  $SW_1 = \pm 125$  Hz;  $SW_2 = 4\,000$  Hz;  $NS = 144$ ;  $T = 303$  K; recycle delay: 2.5 s; data matrix: 2 K  $\times$  128.

### <sup>1</sup>H–<sup>13</sup>C-Shift Correlation

Pulse sequence: D  $1-\pi/2(\text{PH } 1, ^1\text{H})-t_1/2-\pi(\text{PH } 3, ^{13}\text{C})-t_1/2-\Delta_1-\pi/2(\text{PH } 2, ^1\text{H})-\pi/2(\text{PH } 4, ^{13}\text{C})-\Delta_2$ -FID (BB);  $SW_1 = \pm 500$  to  $\pm 900$  Hz;  $SW_2 = 3\,000$  to  $8\,000$  Hz;  $NS = 160$ – $640$ ;  $T = 303$  K;  $PW(\pi/2, ^{13}\text{C}) = 18\ \mu\text{s}$ ;  $PW(\pi/2, ^1\text{H}) = 17\ \mu\text{s}$ ; recycle delay: 2.5 s; data matrix: 2 K (4 K)  $\times$  128 (256);  $\Delta_1 = 4$  ms;  $\Delta_2 = 2.67$  ms.

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